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# Acclimation response of spring wheat in a free-air CO<sub>2</sub> enrichment (FACE) atmosphere with variable soil nitrogen regimes. 1. Leaf position and phenology determine acclimation response

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#### **Abstract**

We have examined the photosynthetic acclimation of wheat leaves grown at an elevated  $CO_2$  concentration, and ample and limiting N supplies, within a field experiment using free-air  $CO_2$  enrichment (FACE). To understand how leaf age and developmental stage affected any acclimation response, measurements were made on a vertical profile of leaves every week from tillering until maturity. The response of assimilation (A) to internal  $CO_2$  concentration ( $C_1$ ) was used to estimate the *in vivo* carboxylation capacity ( $Vc_{max}$ ) and maximum rate of ribulose-1,5-bisphosphate limited photosynthesis ( $A_{sat}$ ). The total activity of ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco), and leaf content of Rubisco and the Light Harvesting Chlorophyll a/b protein associated with Photosystem II (LHC II), were determined. Elevated  $CO_2$  did not alter  $Vc_{max}$  in the flag leaf at either low or high N. In the older shaded leaves lower in the canopy, acclimatory decline in  $Vc_{max}$  and  $A_{sat}$  was observed, and was found to correlate with reduced Rubisco activity and content. The dependency of acclimation on N supply was different at each developmental stage. With adequate N supply, acclimation to elevated  $CO_2$  was also accompanied by an increased LHC II/Rubisco ratio. At low N supply, contents of Rubisco and LHC II were reduced in all leaves, although an increased LHC II/Rubisco ratio under elevated  $CO_2$  was still observed. These results underscore the importance of leaf position, leaf age and crop developmental stage in understanding the acclimation of photosynthesis to elevated  $CO_2$  and nutrient stress.

Abbreviations:  $A_{\text{sat}} - \text{CO}_2$  and light saturated rate of  $\text{CO}_2$ -uptake per unit leaf area;  $\text{CL}-\text{control CO}_2$  and limiting nitrogen supply;  $\text{FL}-\text{elevated CO}_2$  and limiting nitrogen supply;  $\text{CH}-\text{control CO}_2$  and high nitrogen supply;  $\text{FH}-\text{elevated CO}_2$  and high nitrogen supply; IRGA-Infra-red gas analyzer; LHC II-light harvesting chlorophyll a/b protein primarily associated with Photosystem II; RuBP-ribulose-1,5-bisphosphate;  $Vc_{\text{max}}-\text{maximum ribulose-1,5-bisphosphate}$  associated rate of carboxylation in vivo

# Introduction

The concentration of atmospheric CO<sub>2</sub> is expected to double during the 21st century (McElroy 1994). CO<sub>2</sub> concentration is limiting for C<sub>3</sub> photosynthesis, and so

it is generally expected that a rise in atmospheric CO<sub>2</sub> concentration will stimulate photosynthesis and increase dry matter production (Kramer 1981). This has been demonstrated in a wide range of C<sub>3</sub> plants (Kimball 1983; Gifford 1988; Ziska et al. 1991). Increased

CO<sub>2</sub> uptake in C<sub>3</sub> photosynthesis is primarily due to the fact that the increased substrate concentration at the active site of the primary carboxylating enzyme, Rubisco, suppresses photorespiration (Bowes 1996; Drake et al. 1997). Prolonged exposure to elevated CO<sub>2</sub>, especially under nutrient limiting conditions, may result in an acclimatory response of C<sub>3</sub> photosynthesis (Kriedemann and Wong 1984; Sage et al. 1989). Such an acclimatory response may involve a decrease in the amount and activity of Rubisco (Wong 1979; Sage et al. 1989, 1994; Oosten et al. 1992; McKee and Woodward 1994; McKee et al. 1995). Rubisco can account for as much as 50% of the leaf soluble protein and 10-25% of leaf N and is in excess of the level required for photosynthesis when N supply is adequate (Theobald et al. 1998). A decrease in the amount and activity of Rubisco has been observed in wheat grown at elevated CO<sub>2</sub>. With adequate N supply, this decrease does not always completely nullify the increased photosynthesis rate due to elevated CO<sub>2</sub>. When N supply is limiting, it is possible that Rubisco content will be reduced to a level that will lower light saturated photosynthesis, A, and  $A_{\text{max}}$  (Evans and Farquhar 1991; Webber et al. 1994). However, results of experiments studying the interactions of CO<sub>2</sub> enrichment and N nutrition have been inconclusive.

While several studies have reported a decrease in leaf Rubisco content with growth at elevated CO<sub>2</sub>, others have found no significant loss of this protein, or only a decrease in its activation (reviewed by Long and Drake 1992; Sage 1994; Webber et al. 1994). Wong (1979) showed, in a short-term study with cotton, that down-regulation because of elevated CO2 was larger at low N-supply than at higher N-supply. Oberbauer et al. (1986) also found in a 3-month study that CO<sub>2</sub> enrichment of Ledum resulted in a larger down-regulation in plants grown with lower N supply. On the other hand, there was no evidence for down-regulation in another short-term study on cotton by Wong (1990), or in a short-term study of Phaseolus vulgaris grown at highand low-N supply (Radoglou et al. 1992). Although part of this variation may result from an artificial limitation of rooting volume (Arp 1991), variation in decrease in Rubisco content was still evident among studies in which care was taken to provide a large rooting volume (Long and Drake 1992). Even within the same species, for example wheat, some studies have reported, or inferred, a decrease in Rubisco with growth in elevated CO<sub>2</sub> concentrations (McKee and Woodward 1994) whilst others have not (Delgado et al. 1993; Farage et al. 1998). In a long-term glasshouse

study of winter wheat grown at two levels of CO<sub>2</sub> and two levels of N encompassing the full growing season, no evidence of down-regulation was found (Mitchell et al. 1993), even though the plants were grown in pots. This may be attributed to the ability of cereal crops to respond to increased assimilate supply during the vegetative stage by increasing tillering, and during the reproductive phase by translocating more assimilate to the ear (Petterson and McDonald 1994)

Growth of plants under Free-air CO<sub>2</sub> enrichment (FACE) allows a direct study of the effect of elevated CO2 under field conditions without disturbances frequently encountered within controlled environment and open-top chambers (Hendrey et al. 1993; McLeod and Long 1999). Additionally, FACE studies allow the effects of elevated CO2 to be monitored continuously throughout a growing season. In an earlier study, spring wheat growing in FACE and well-watered conditions showed reductions in carboxylation efficiency and Rubisco content due to CO<sub>2</sub> enrichment in the top three fully-expanded leaves (Nie et al. 1995; Osborne et al. 1998). Comparing measurements of different growth stages made in separate years, photosynthetic acclimation was observed at the grain filling stage, and was more pronounced in lower leaves when compared to the uppermost fully expanded leaf (Osborne et al. 1998). However, interactions of CO<sub>2</sub> enrichment and nutrient stress were not addressed.

Based upon the previous discussion, there are a number of factors that may lead to an acclimation response to elevated CO<sub>2</sub> in the field. In particular, periods of source sink imbalance, especially when reproductive sinks are developing, may be important determinants for acclimation. Therefore, it is critical that the interaction between leaf age, crop phenology and photosynthetic acclimation in response to growth in elevated CO<sub>2</sub> is studied in more detail. We have tested the hypothesis that leaf age, phenology and Nsupply will be important in determining the response of a wheat crop to elevated CO2 by studying the interaction of CO2 enrichment and two levels of N nutrition on acclimation of photosynthesis in a fieldgrown spring wheat crop, comparing the upper-most fully expanded leaf and lower canopy leaves, at each successive developmental stage from tillering through grainfill.

#### Materials and methods

An experiment was conducted in 1997 to determine in-

teractive effects of elevated CO2 and limited soil nitrogen on spring wheat (Triticum aestivum L. cv. Yecora Rojo) at the University of Arizona Maricopa Agricultural Center (MAC), Maricopa, Arizona, USA. The free-air CO<sub>2</sub> enrichment (FACE) technique was used to enrich the air in circular plots within a wheat field similar to prior experiments (Hendrey et al. 1993; Wall and Kimball, 1993; Dugas and Pinter 1994; Wechsung et al. 1995; Kimball et al. 1995, 1999). For this experiment, the FACE plots were enriched by 200  $\mu$ mol/mol above ambient ( $\sim 360 \,\mu\text{mol/mol}$ ). Unlike the prior experiments, air blowers were installed in the non-CO<sub>2</sub> enriched ambient control plots to provide similar air movement to the FACE plots (Kimball et al. 1999). The FACE treatment was applied continuously from leaf emergence to grain harvest. Each of the circular plots was split into semicircular halves, with each half receiving either an ample (350 kg ha<sup>-1</sup> season<sup>-1</sup>) or a limiting (15 kg ha<sup>-1</sup> season<sup>-1</sup>) level of nitrogen fertilizer.

#### Gas-exchange measurements

Measurements were made on leaf material obtained from an undisturbed portion of the canopy. Plant material was sampled prior to 7:30 am to avoid any effects of photoinhibition or water deficit on the leaves. Plants were placed in sealed plastic bags and stored at 10 °C in darkness until measurements were made, according to procedures described by Osborne et al. (1998). The leaf to be measured was excised at the ligule under water and placed in the cuvette of a LI-6400 (LiCor, Inc. Lincoln, Nebraska) with the [CO<sub>2</sub>] set at the growth [CO<sub>2</sub>]. The leaf was positioned in the chamber at an equal distance from the tip and the ligule. The chamber was illuminated at 1200  $\mu$ mol photons  $m^{-2}$  s<sup>-1</sup> and the leaf temperature maintained at  $20 \pm 0.1$  °C. Relative humidity was maintained at 50%. The LI-6400 software/graphics package was used to determine whether steady-state photosynthesis had been reached. When steady-state photosynthesis was reached, the program to vary intercellular [CO<sub>2</sub>] (C<sub>i</sub>) was used to measure photosynthesis at C<sub>i</sub>'s ranging from 50 to 1000  $\mu$ mol CO<sub>2</sub> mol<sup>-1</sup>. The initial linear slope of the A/Ci response was used to estimate Vcmax according to the method of Wullschleger (1993) using the parameter coefficients of Harley et al. (1992).  $A_{\text{sat}}$  was measured at 1000  $\mu$ mol CO<sub>2</sub> mol<sup>-1</sup>. At the end of the A/Ci measurement program, the leaf was allowed to reach steady state photosynthesis at its growth [CO<sub>2</sub>]. When steady-state photosynthesis was reached, the leaf was removed from the cuvette and immediately freeze-clamped with a liquid nitrogen-cooled clamp, and stored in liquid nitrogen until biochemical assays were conducted.

#### Biochemical assays

Leaf tissue was removed from liquid nitrogen and ground in an ice-cold glass homogenizer containing 100 mM Tricine (pH 8.0), 5 mM MgCl<sub>2</sub>, 0.1 mM EDTA, 5 mM DTT, 1% (w/v) PVP, 0.05% (v/v) Triton X-100 and 1 mM phenylmethylsulfonyl fluoride at a ratio of 1 cm<sup>2</sup> leaf tissue to 1 ml buffer. An aliquot was assayed for full activity and in situ activity of Rubisco following the assay procedure of Salvucci and Anderson (1987), with the exception that casein was not included in the assay mixture. Aliquots were saved for chlorophyll and total soluble protein determination and stored at -20 °C. Chlorophyll concentrations were determined according to Arnon (1949). Additional aliquots were boiled in SDS sample buffer and used for SDS-PAGE as described by Nie et al. (1995). Following SDS-PAGE, the gels were stained with coomassie blue and the relative intensity of each band quantitated using a laser scanning densitometer. The relative amount of Rubisco and LHC II was determined from the area under the corresponding peaks as described previously (Nie et al. 1995).

### Statistical analysis

The effect of nitrogen on photosynthetic acclimation to elevated  $CO_2$  was examined using a two way ANOVA for  $A_{\text{sat}}$ ,  $V_{\text{cmax}}$  and Rubisco activity. Gas exchange measurements and biochemical assays were replicated three times for each sample treatment.

## Results

The A/C<sub>i</sub> response of individual leaves of FACE grown spring wheat was measured throughout the growing season for the uppermost, fully expanded leaf (flag leaf) and next two successively older leaves (flag-1 and flag-2). Material was harvested from each leaf for biochemical analysis (see 'Materials and methods'). The results describe first a statistical analysis of the overall effect of elevated CO<sub>2</sub> and N supply on photosynthetic parameters. This is followed by a more detailed analysis of these parameters for each leaf at different growth stages, to understand

Table 1. Overall analysis of season long pooled data for each leaf showing the effects of ambient and high CO<sub>2</sub> at both N levels

Leaf	Low CO <sub>2</sub>	High CO <sub>2</sub> <sup>a</sup>	P > F	Low N	High N	P > F	Lo	Low N	HI	High N	Interaction
							Low $CO_2$	High CO <sub>2</sub> <sup>b</sup>	LowCO <sub>2</sub>	${ m HighCO_2}^{ m b}$	P > F
Vcmax Flag $(\mu \text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1})$	8.44.8	42.1 (6) <sup>b</sup>	0.0393	41.1 (10.3)	45.8	0.0020	43.6	38.6 (11.5)	46.05	45.6 (1.0)	0.1209
Flag-1	41.4	30.3 (26.8)	0.0001	31.8 (20.3)	39.9	0.0373	35.7	27.9 (21.8)	47.1	32.7	0.0997
Flag-2	29.7	23 (22.6)	0.1783	21.4 (31.6)	31.3	0.0001	24.2	18.5 (23.6)	35.2	27.4 (22.2)	0.5269
$A_{\text{sat}}$ Flag $(\mu \text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1})$	27.4	26.5	0.3709	25.7	28.2	0.1339	26.2	25.2 (3.8)	28.6	27.9 (2.4)	0.8234
Flag-1	22.1	19.2	0.0792	18.6 (18.1)	22.7	0.0662	19.5	17.8	24.6	20.7 (15.9)	0.3677
Flag-2	15	12.4 (17.3)	0.0116	11.5 (27.7)	15.9	0.0001	12.4	10.6 (14.5)	17.6	14.2 (19.3)	0.4356
Total Chl. Flag (mg Chl cm <sup>-2</sup> )	41.2	40 (2.9)	0.1666	32.3	48.9	0.0158	32.9	31.6 (4.0)	49.62	48.3	0.9842
Flag-1	41.6	39.2	0.2715	30.2 (40.3)	50.6	0.0017	31.1	29.3	52	49.1	0.6773
Flag-2	33.1	30.8	0.2776	22.1 (47.1)	41.8	0.0024	23.5	20.8	42.7	40.9	0.8237
Rubisco Flag $(\mu \text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1})$	75.6	66 (12.7)	0.0001	54.7 (37.1)	86.9	0.0012	59.3	50.2 (15.3)	92	81.8 (11.1)	0.8029
Flag-1	59	44.6 (24.4)	0.3967	39 (39.6)	64.6	9660.0	45.1	32.9	72.9	56.3 (22.8)	0.5973
Flag-2	32	21.7 (32.2)	0.0018	19.2 (44.5)	34.6	0.0001	23.1	15.2 (34.2)	41	28.2 (31.2)	0.4419

<sup>a</sup>Value in parentheses denotes percent decrease from ambient CO<sub>2</sub>.

<sup>b</sup>Value in parentheses denotes percent decrease from high N.

how any response was influenced by leaf position and developmental age of the crop.

### Overall treatment effect

Table 1 summarizes an overall analysis, by leaf, of season-long pooled data. A two-way ANOVA indicates a decrease in  $Vc_{\text{max}}$  and  $A_{\text{sat}}$  due to  $CO_2$  enrichment or N stress for the flag, flag-1 and flag-2 leaf. This decrease in  $Vc_{\text{max}}$  and  $A_{\text{sat}}$  was more pronounced for leaves lower in the canopy, indicating a significant influence of leaf age and position on the parameters measured. Significant reduction in total chlorophyll and total Rubisco activity due to the N stress or CO<sub>2</sub> enrichment were also observed, again with the greatest reduction occurring in older leaves (Table 1). CO<sub>2</sub> enrichment resulted in greater reduction at low N than at high N for all parameters in the flag leaf (Table 1). The flag-1 leaf showed greater reduction in  $Vc_{\text{max}}$  and  $A_{\text{sat}}$  due to  $CO_2$  enrichment under high N than under low N. Total chlorophyll and Rubisco activity showed larger reductions under low N in the flag-1 leaf. Reduction due to CO<sub>2</sub> enrichment for the flag-2 leaf was greater under low N for Vc<sub>max</sub>, total chlorophyll and Rubisco activity, and greater under high N for  $A_{sat}$ .

#### Analysis by developmental stage

Photosynthetic gas exchange parameters were measured every week from tillering, which commenced 72 days after planting (DAP 72), until maturity at DAP 143. Results from this analysis are presented in Figures 1 and 2. Prior to canopy closure at the boot stage, only the uppermost fully expanded leaf was measured. Photosynthetic gas exchange parameters (Figures 1 and 2) and Rubisco activity (Figure 3) progressively decreased with leaf position down the stem. Comparing leaves grown at ambient CO<sub>2</sub> and high N, A<sub>sat</sub> and Vc<sub>max</sub> decreased by approx 25% in the flag-2 leaf compared to the flag leaf at the heading stage. By the late milk dough stage, this reduction was 45%. Rubisco activity (Figure 3) was reduced 50% in the flag-2 leaf at the late milk dough stage. At the heading stage, the chlorophyll a/b ratio (not shown) was reduced from approximately 3.2 in the flag leaf, to 2.7 in the flag-2 leaf, consistent with a slight increase in content of the chlorophyll a/b-containing light harvesting complexes in response to the lower light level deeper in the canopy. At the late milkdough stage, the chlorophyll a/b ratio was 2.9 in the flag leaf and 2.5 in the flag-2 leaf.

#### Flag leaf

A gradual decline in A<sub>sat</sub> was observed from an average value of 40  $\mu \mathrm{mol}~\mathrm{m}^{-2}~\mathrm{s}^{-1}$  at tillering to approx 17  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup> at the soft dough stage (Figure 1). Neither elevated CO<sub>2</sub> nor N-stress had any pronounced effect on  $A_{\text{sat}}$  at any growth stage. A similar progressive decline in Vc<sub>max</sub> was observed during the growing season (Figure 2). Significant  $CO_2 \times N$  interactions on Vc<sub>max</sub> were not observed at any growth stage. Rubisco full activity (Figure 3) was reduced by N-stress, ranging from an approximately 23% reduction at stem extension, to a 46% reduction by the end of anthesis. Significant CO2 × N interactions for Rubisco activity were found at several different growth stages. At tillering, milkdough stage (DAP122) and late milkdough stage (DAP 126) Rubisco activity was reduced 21% (P = 0.0287), 46% (P = 0.0192), 35% (P = 0.2774), respectively, by CO<sub>2</sub> enrichment and Nstress. At late anthesis (DAP 117), Rubisco activity was reduced 25% (P = 0.3244) by CO<sub>2</sub> enrichment under low N, and 36% (P = 0.0908) under high N. Total chlorophyll content (Figure 4) was reduced 47% by N-stress (P = 0.0155) at mid-anthesis, and a similar Nstress reduction in total chlorophyll was observed the remainder of the growing season.

### Flag-1

 $A_{\text{sat}}$  (Figure 1) was reduced 33% by CO<sub>2</sub> enrichment at both low (P = 0.0165) and high N (P = 0.0109) at the heading stage (DAP 99). At the early milk-dough stage (DAP 122), CO<sub>2</sub> enrichment reduced A<sub>sat</sub> (41%, P = 0.0042) at both N levels. Significant  $CO_2 \times N$ interactions in  $A_{\text{sat}}$  were not observed. The highest rates of Vc<sub>max</sub> (Figure 2) and Rubisco activity (Figure 3) were observed for plants grown at atmospheric CO<sub>2</sub> and high N. Largest reductions in Vc<sub>max</sub> due to elevated CO<sub>2</sub> were observed under high N at early anthesis, mid-anthesis and soft dough stage.  $CO_2 \times N$ interactions were also observed in Rubisco activity during boot stage (DAP 92) when elevated CO<sub>2</sub> resulted in a 28% reduction in Rubisco activity (P = 0.0581) under low N, but had no effect under high N (Figure 3). From mid-anthesis onward,  $CO_2 \times N$  interactions in Rubisco activity were due to reduced activity at elevated CO<sub>2</sub> and high N (reductions of 36%, 34% and 18% at DAP 112, 117 and 126, respectively). By the heading stage, N-stress had a clear effect on total chlorophyll content (Figure 4). During early anthesis (DAP 108) and onwards, total chlorophyll was re-

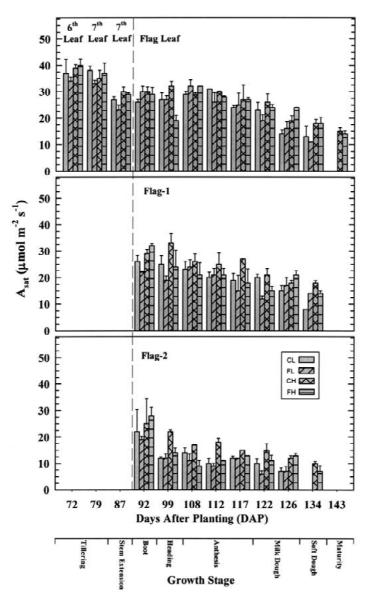


Figure 1.  $A_{sat}$  for the flag, flag-1 and flag-2 leaves in wheat grown in elevated  $CO_2$  and in control  $CO_2$  conditions with ample and limiting N supplies, measured at different developmental stages. The days after planting (DAP) and growth stage, determined by Zadok's scale, are indicated beneath the figure. CL-control  $CO_2$  and limiting nitrogen supply; FL-elevated  $CO_2$  and limiting nitrogen supply; FL-elevated FL-control FL-elevated FL-control FL-elevated FL-

duced by 40–48% by N stress. No significant changes were observed in the chlorophyll a/b ratio.

#### Flag-2

Significant  $CO_2 \times N$  interactions were found in  $A_{sat}$  for the flag-2 leaf at the heading, early and midanthesis stages, and milkdough stages (Figure 1). At heading, early and mid-anthesis,  $CO_2$  enrichment at high N reduced Asat by 29%, 42% and 47%, re-

spectively. At the early milk dough stage,  $A_{\rm sat}$  was decreased by 40% at low N and 27% at high N. The highest rates of Rubisco activity and  $Vc_{\rm max}$  were observed for the plants grown at atmospheric CO<sub>2</sub> and high N, similar to that observed for the flag-1 leaf.  $Vc_{\rm max}$  was reduced (44%) by N-stress at both control and elevated CO<sub>2</sub> levels at early anthesis and early milk-dough stages (Figure 2). At heading, CO<sub>2</sub> enrichment reduced  $Vc_{\rm max}$  at high N (33%; P=0.0186).

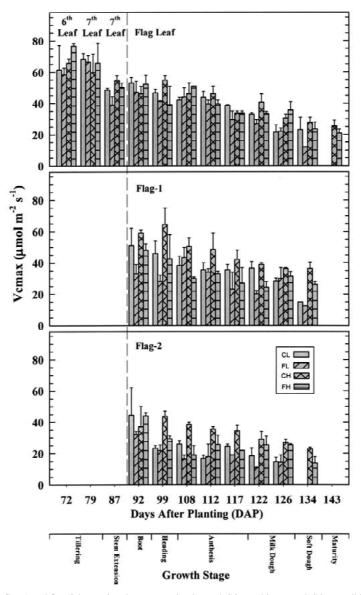


Figure 2.  $Vc_{max}$  for the flag, flag-1 and flag-2 leaves in wheat grown in elevated  $CO_2$  and in control  $CO_2$  conditions with ample and limiting N supplies, measured at different developmental stages (Figure 1).

At late anthesis,  $Vc_{\text{max}}$  was reduced 22% (P = 0.1477) by CO<sub>2</sub> enrichment at low N, and 36% at high N (P = 0.0110). At the early milk dough stage,  $Vc_{\text{max}}$  was reduced more at low N (42%) than at high N (12%).

Rubisco activity (Figure 3) showed significant  $CO_2 \times N$  interactions at many growth stages. At the boot stage, elevated  $CO_2$  caused a 43% reduction in Rubisco activity at low N (P=0.3729). At the three anthesis stages,  $CO_2$  enrichment at high N reduced Rubisco activity by an average of 55%. At the early milk dough stage, this pattern reversed, and Rubisco

activity was reduced 63% at low N and 27% at high N.

At the heading stage, N-stress reduced total chlorophyll (45%, P = 0.0177; Figure 4). The N-stressed reduction in total chlorophyll progressively increased at subsequent growth stages. By late milk dough stage, total chlorophyll was decreased 69% (P = 0.0424) by N stress. The chlorophyll a/b ratio of 2.5 did not significantly change at any of the different growth stages (not shown).

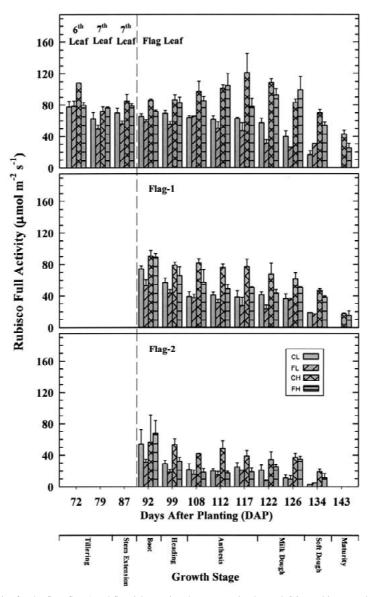


Figure 3. Rubisco full activity for the flag, flag-1 and flag-2 leaves in wheat grown in elevated  $CO_2$  and in control  $CO_2$  conditions with ample and limiting N supplies, measured at different developmental stages (Figure 1). Leaf material was sampled from the same portion of the leaf for which  $A_{\text{sat}}$  and  $Vc_{\text{max}}$  were determined.

#### Protein content

At selected growth stages, total protein was isolated from leaf material and size fractionated by SDS–PAGE (Figures 5 and 6). Rubisco content of the flag-2 leaf, at normal N, was reduced 40–50% compared to the flag leaf. Changes in the accumulation of Rubisco protein were found to correlate closely with the observed changes in total Rubisco activity. This indicated that the decreased  $Vc_{\rm max}$  measured *in situ* was due

to reduced Rubisco activity, resulting from a lowered Rubisco content per leaf surface area. LHC II content under control conditions (high N and atmospheric CO<sub>2</sub> level) increased 25% from DAP 92 to DAP 126 (Figures 5 and 6). The flag-1 and flag-2 leaves had a 30–50% higher LHC II content at DAP 122 and 126 compared with the flag leaf at DAP 92. The net result of changes in LHC II – Rubisco content was an increase in the ratio of LHC II to Rubisco in the older leaves, relative to the flag leaf. Elevated CO<sub>2</sub> also in-

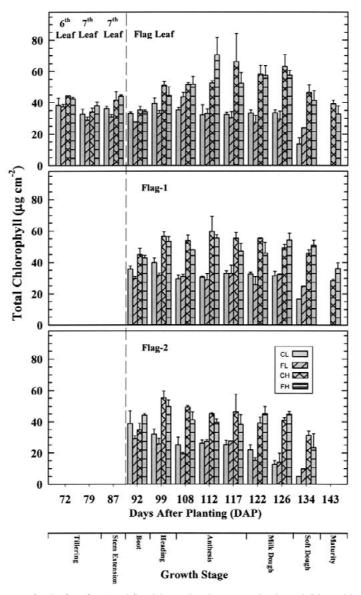


Figure 4. Total chlorophyll content for the flag, flag-1 and flag-2 leaves in wheat grown in elevated  $CO_2$  and in control  $CO_2$  conditions with ample and limiting N supplies, measured at different developmental stages.

creased the ratio of LHC II to Rubisco in the flag-1 and flag-2 leaves under ample N conditions. Under low N, the LHC II content and Rubisco content was reduced 50--75% in the flag-2 leaf, indicating an effect of N stress on the accumulation of these two proteins. Nevertheless, the LHC II–Rubisco ratio still increased in the older leaves under the low N conditions, although the effect of elevated  $CO_2$  was less.

### Discussion

The acclimatory response of individual leaves to growth under N-stress and in elevated  $CO_2$  was strongly dependent upon leaf position in the canopy and upon the developmental stage of the crop. A progressive decline in  $Vc_{max}$  was observed with age of the flag leaf, but there was no significant acclimatory response to elevated  $CO_2$  at either N-level. The flag-1 and flag-2 leaves showed a significant acclimation of  $Vc_{max}$  in response to growth in elevated  $CO_2$ . De-

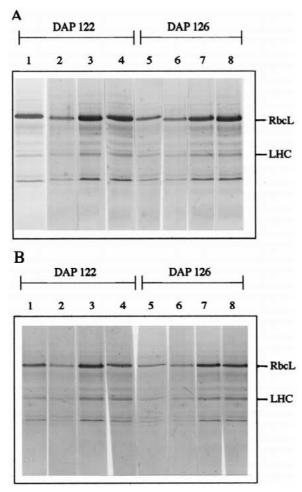


Figure 5. Protein profile of the flag (A) and flag-2 (B) leaves of wheat grown in elevated CO<sub>2</sub> and in control CO<sub>2</sub> conditions with ample and limiting N supplies, measured at different developmental stages (DAP 122 and DAP 126). Protein isolates were size fractionated by SDS–PAGE, and polypeptides visualized by staining with Coomassie blue. Lanes 1 and 5, CL; 2 and 6, FL; 3 and 7, CH; 4 and 8, FH.

pending on the developmental stage, this acclimation occurred under either N-stress, ample N or under both N levels. Acclimation of  $Vc_{max}$  in the lower leaves was apparent at the heading stage through to the early milk dough stage, but was not observed at the late milk dough stage. In an earlier study of FACE-grown wheat, comparing measurements of different growth stages made in separate years, photosynthetic acclimation in lower leaves at the grain filling stage was also observed (Osborne et al. 1998). In this study, a season-long analysis involving measurements at eight different growth stages clearly indicates the importance of leaf position and phenology in determining the

response of individual leaves to elevated CO<sub>2</sub>. Furthermore, the interaction of N-supply with growth in elevated CO<sub>2</sub> also is dependent on which leaf is studied and on the developmental age of the crop at the time of measurement.

Rubisco can account for as much as 50% of the leaf soluble protein and is a major N investment of the plant. Rubisco can accumulate in excess of that required for photosynthesis when N supply is adequate. However, when N-supply is limiting, it is expected that Rubisco accumulation will be reduced to a level sufficient for maximum photosynthesis, freeing up N for use elsewhere in the plant (Evans and Farquhar 1991; Webber et al. 1994). Experimental evidence has shown that a decline in Vc<sub>max</sub> is due to a decreased Rubisco content or activity. Based on the aforementioned considerations, reduction in Vc<sub>max</sub> with elevated CO<sub>2</sub> is expected to be more pronounced under N-stress growth conditions. In the flag leaf, the progressive decline in Vcmax with leaf age was not paralleled by a similar decline in Rubisco activity at high N. Osborne et al. (1998) attributed this to a decreased activation state of Rubisco with leaf age. The reduced Rubisco activity at low N at each developmental stage, however, was matched by a corresponding decrease in Vc<sub>max</sub>. Neither Vc<sub>max</sub> nor Rubisco activity in the unshaded flag leaf was significantly altered by elevated CO<sub>2</sub>, consistent with earlier FACE studies of wheat (Nie et al., 1995). In lower leaves (flag-1 and flag-2), the decline in  $Vc_{\text{max}}$  with age and developmental stage correlated with reduced Rubisco activity and content. The decline in Vc<sub>max</sub> due to elevated CO<sub>2</sub> was more pronounced when N supply was adequate, particularly during anthesis (Figure 2). A decrease in Vc<sub>max</sub> and Rubisco activity for N-stressed leaves in elevated CO<sub>2</sub> was only observed prior to anthesis and again during late anthesis and early milk dough stages. In lower leaves, elevated CO<sub>2</sub> had little effect on the level of leaf total soluble protein (not shown), especially at low N. This would imply that acclimation observed under these conditions was specific for Rubisco, and was not a dilution effect.

In monocotyledons, such as wheat, new leaves emerge into full sunlight at the top of the canopy and then become more shaded as new leaves emerge above them. The decline in  $Vc_{\rm max}$  and  $A_{\rm sat}$  with depth in the canopy, therefore, corresponds with the degree of shading and age of the leaves. Typical shade responses of the photosynthetic apparatus include an increase in LHC II proteins and a decrease in Rubisco content (Anderson 1986; Evans 1993, 1996). In this study,

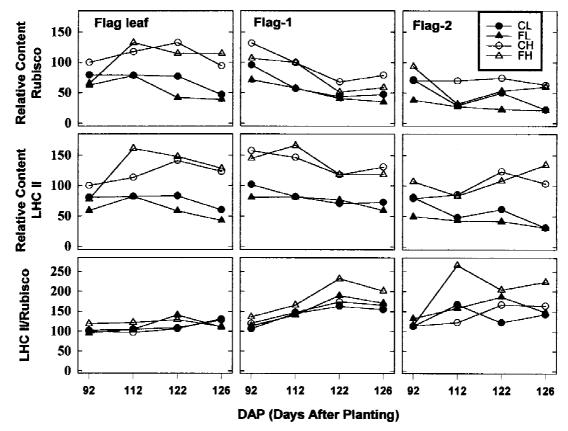


Figure 6. Rubisco, LHC II and LHC II/Rubisco for the flag, flag-1 and flag-2 leaves in wheat grown in elevated CO<sub>2</sub> and in control CO<sub>2</sub> conditions with ample and limiting N supplies, measured at different developmental stages. The relative amount of Rubisco and LHC II was determined by scanning of gels shown in Figure 5 (see 'Materials and methods'). Values are the protein content determined on a leaf area basis, and expressed in arbitrary units relative to the absorption of the corresponding protein in control leaves at DAP 92.

shade adaptation was reflected by the increased LHC II/Rubisco ratio observed in the lower leaves. With ample N, LHC II content was not reduced in the lower leaves, indicating that the increased ratio of LHC II - Rubisco was related to light acclimation and not senescence. Under limiting N supply, the LHC II content was reduced in the flag-2 leaf, suggesting that some senescence may have occurred. Even with limiting N supply, the LHC II - Rubisco ratio increased in the flag-2 leaf indicating light acclimation. Canopy architecture and light penetration, however, were not significantly changed by growth in elevated CO<sub>2</sub> when N supply was adequate. Under low N, canopy architecture was significantly different to that of plants growing at ample N (Brooks et al. 2000). At low N, the canopy was more erectophile, but this resulted in only a slight increase in light penetration to the lower leaves compared to ample N conditions, and was unaffected by elevated CO2. This was reflected by the

fact that the LHC II – Rubisco ratio in the N-stressed leaves was similar to that of leaves of plants receiving adequate N (Figure 6). Therefore, it is unlikely that the acclimation response to elevated  $\rm CO_2$  observed in the lower leaves was due to increased shading in either the N-stress or ample N treatments.

Several factors may contribute to the different responses of wheat leaves at different developmental stages in elevated  $CO_2$ . Changes in resource allocation will occur as the crop switches from vegetative growth to the production of reproductive structures and grain filling. During the developmental sequence, leaves will transition from an initial sink to a source of photosynthate and N. In particular, N tied up in Rubisco is likely to be reallocated to grain development (Simpson et al. 1983; Simpson, 1992), and such a reassignment would explain the more pronounced decline in  $Vc_{\rm max}$  and Rubisco activity at elevated  $CO_2$  in leaves lower in the canopy at the late anthesis and

grain filling stage. In addition, growth in elevated  $CO_2$  with ample N could result in increased growth and rate of production of the reproductive structures compared to growth at either  $CO_2$  concentration under N-stress. This increased growth would explain the observed dilution of leaf N content (Sinclair et al. 1998), particularly in leaves of plants grown at low N and elevated  $CO_2$  and lead to an acclimatory decline in  $Vc_{max}$  and  $A_{sat}$  (Osbourne et al. 1998). In any case, this acclimation response would be expected to be functionally significant because less Rubisco would be required to maintain a constant photosynthetic rate in elevated  $CO_2$  conditions.

Results from *in situ* studies also reported in this issue indicate as much as a 30% increase in carbon gain in the uppermost fully expanded leaves due to elevated Ca (Wall et al. 2000). Some stimulation of  $C_3$  photosynthesis at elevated  $CO_2$  is expected despite the observed reduction in  $A_{\rm sat}$  and  $V_{\rm cmax}$  because the oxygenation of RuBP is suppressed. This is in agreement with results reported by Osborne et al. (1998), who interpreted this to indicate that acclimation was best interpreted as increased efficiency of resource allocation rather than an adverse reaction to elevated  $CO_2$ .

In contrast, it is important to emphasize that the whole-canopy net assimilation rate showed at most only a 10% increase (Brooks et al. 2000). Presumably there are compensatory changes in canopy architecture that minimize the overall effect of the responses observed at the individual leaf level. Indeed, results of above and below ground biomass production were in better agreement with the whole-canopy net assimilation rates than to results obtained from individual leaves.

In conclusion, there is still considerable controversy as to the extent to which photosynthetic apparatus will acclimate in response to future elevated CO<sub>2</sub> atmospheres. This is partly due to the fact that previous work has focused only on the uppermost fully expanded leaf, and reletively few studies (e.g. Osborne et al. 1998) have considered leaf age and crop phenology as a variable. This work demonstrates that acclimation of the photosynthetic apparatus of wheat will occur in response to elevated CO<sub>2</sub>. However, the response is most pronounced in lower shaded leaves and at later stages of crop development. Taken together with results reported in other papers in this volume (Brooks et al. 2000; Wall et al. 2000), this work emphasizes the inherent complexities in trying to predict

the acclimatory responses of a wheat crop to growth at elevated CO<sub>2</sub>.

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#### References

- Anderson JM (1986) Photoregulation of the composition, function and structure of thylakoid membranes. Ann Rev Plant Physiol 37: 93–136
- Arp WJ (1991) Effects of source-sink relations on photosynthetic acclimation to elevated CO<sub>2</sub>. Plant Cell Environ 14: 869–876
- Bowes G (1996) Photosynthetic responses to changing atmospheric carbon dioxide concentrations. In: Baker NA (ed) Photosynthesis and the Environment, pp 387–407. Kluwer Academic Publishers, Dordrecht, The Netherlands
- Brooks TJ, Wall GW, Pinter PJ Jr, Kimball BA, LaMorte RL, Leavitt SW, Matthias AD, Adamsen FJ, Hunsaker DJ and Webber AN (2000) Acclimation response of spring wheat in a free-air CO<sub>2</sub> enrichment (FACE) atmosphere with variable soil nitrogen regimes. 3. Canopy architecture and gas exchange. Photosynthesis Research 66: 97–108 (this issue)
- Delgado E, Mitchell RAC, Parry MAJ, Driscoll SP, Mitchell VJ and Lawlor DW (1994) Interacting effects of CO<sub>2</sub> concentration, temperature and nitrogen supply on photosynthesis and composition of winter wheat leaves. Plant Cell Environ 17: 1205–1213
- Drake BG, Gonzalez-Meler MA, Long SP (1997) More efficient plants: A consequence of rising atmospheric CO<sub>2</sub>? Ann Rev Plant Physiol Plant Mol Biol 48: 609–639
- Dugas WA and Pinter PJ Jr (eds) 1994. The Free-Air Carbon Dioxide Enrichment (FACE) Cotton Project: A New Field Approach to Assess the Biological Consequences of Global Change. Agri For Meteorol 70: 1–342
- Evans JR (1993) Photosynthetic acclimation and nitrogen partitioning within a lucerne canopy. I. Canopy characteristics. Aust J Plant Physiol 20: 55–67
- Evans JR and Farquhar GD (1991). Modeling canopy photosynthesis from the biochemistry of the C<sub>3</sub> chloroplast. In: Boote KJ

- and Loomis RS (eds) Modeling Crop Photosynthesis from Biochemistry to Canopy, pp 1–16. Crop Science Society of America, Inc., Madison, Wisconsin
- Farrage PK, McKee IF and Long SP (1998) Does a low nitrogen supply necessarily lead to acclimation of photosynthesis to elevated CO<sub>2</sub>? Plant Physiol 118: 573–580
- Harley PC, Thomas RB, Reynolds JF and Strain BR (1992) Modelling photosynthesis of cotton grown in elevated CO<sub>2</sub>. Plant Cell Environ 15: 271–282
- Hendrey GR (ed) (1993) Free-Cir Carbon Dioxide Enrichment for Plant Research in the Field. C.K. Smoley, Boca Raton, Florida, 308 pp
- Kramer PG (1981) Carbon dioxide concentration, photosynthesis and dry matter production. Bioscience 20: 1201–1208
- Kimball BA (1983) Carbon dioxide and agricultural yield: An assemblage and analysis of 430 prior observations. Agron J 75: 779–788
- Kimball BA, Pinter PJ Jr, Garcia RL, LaMorte RL, Wall GW, Hunsaker DJ, Wechsung G, Wechsung F and Kartschall T (1995) Productivity and water use of wheat under free-air CO<sub>2</sub> enrichment. Glob Change Biol 1: 429–442
- Kriedemann PE and Wong SC (1984) Growth response and photosynthetic acclimation to  $CO_2$ : Comparative behavior in two  $C_3$  crop species. Acta Hort 162: 113–120
- Long SP and Drake BG (1992) Photosynthetic CO<sub>2</sub> assimilation and rising atmospheric CO<sub>2</sub> concentrations. In: Baker NR and Thomas H (eds) Crop Photosynthesis: Spatial and Temporal Determinants, pp 69–95. Elsevier Scientific Publishers, Amsterdam
- McElroy MB (1994) Climate of the earth: An overview. Environ Pollution 83: 3–21
- McKee IF and Woodward FI (1994) The effect of growth at elevated CO<sub>2</sub> concentrations on photosynthesis in wheat. Plant Cell Environ 17: 853–859
- McLeod AR and Long SP (1999) Free-air carbon dioxide enrichment (FACE) in global change research: a review. Adv Ecol Res 28: 1–56
- Mitchell RAC, Mitchell VJ, Driscoll SP, Franklin J and Lawlor DW (1993) Effects of increased CO<sub>2</sub> concentration and temperature on growth and yield of winter wheat at two levels of nitrogen application. Plant Cell Environ 16: 521–529
- Nie GY, Long SP, Garcia RL, Kimball BA, LaMorte RL, Pinter PL Jr, Wall GW and Webber AN (1995) Effects of free-air CO<sub>2</sub> enrichment on the development of the photosynthetic apparatus in wheat, as indicated by changes in leaf proteins. Plant Cell Environ 18: 855–864
- Oberbauer SF, Oechel WC and Reichers GH (1986) Soil respiration of Alaskan tundra at elevated atmospheric carbon dioxide concentrations. Plant Soil 96: 145–148
- Osborne CP, LaRoche J, Garcia RL, Kimball BA, Wall GW, Pinter PJ Jr, LaMorte RL, Hendry GR and Long SP (1998) Does leaf position within a canopy affect acclimation of photosynthesis to elevated CO<sub>2</sub>? Analysis of a wheat crop under free-air CO<sub>2</sub> enrichment. Plant Physiol 117: 1037–1045
- Petterson R and McDonald JS (1994) Effects of nitrogen supply on the acclimation of photosynthesis to elevated CO<sub>2</sub>. Photosyntn Res 39: 389–400
- Pinter PJ Jr. Kimball BA, Garcia RL, Wall GW, Hunsaker DJ, and LaMorte RL (1996) Free-air CO<sub>2</sub> enrichment: Responses of

- cotton and wheat crops. In: Mooney HA and Koch GN (eds) Terrestrial Ecosystem Response to Elevated Carbon Dioxide, pp 215–249. Academic Press, Orlando, Florida
- Sage RF (1994) Acclimation of photosynthesis to increasing atmospheric CO<sub>2</sub>: The gas exchange perspective. Photosynth Res 39: 351–368
- Sage RF, Sharkey TD and Seemann JR (1989) Acclimation of photosynthesis to elevated CO<sub>2</sub> in five C<sub>3</sub> species. Plant Physiol 89: 590–596
- Salvucci ME and Anderson JC (1987) Factors affecting the activation state and the level of total activity of Ribulose Bisphosphate Carboxylase in tobacco protoplasts. Plant Physiol 85: 66–71
- Simpson RJ (1992) Carbon and nitrogen budgets within the plant. In Baker NR and Thomas H (eds) Crop Photosynthesis: Spacial and Temporal Determinants, pp 105–129. Elsevier Science Publishers, Amsterdam
- Simpson RJ, Lambers H and Dalling MJ (1983) Nitrogen redistribution during grain growth in wheat (*Triticum aestivum* L.) IV. Development of a quantitative model of the translocation of nitrogen to the grain. Plant Physiol 71: 7–14
- Theobald JC, Mitchell RAC, Parry MAJ and Lawlor DW (1998) Estimating the excess investment in ribuslose-1,5-bisphosphate carboxylase/oxygenase in leaves grown under elevated CO<sub>2</sub>. Plant Physiol 118: 945–955
- Tissue DT, Thomas RB and Strain BR (1993) Long-term effects of elevated CO<sub>2</sub> and nutrients on photosynthesis and Rubisco in loblolly pine. Plant Cell Environ 16: 859–865
- Van Oosten JJ, Afif D and Dizengremel P (1992) Long-term effects of a CO<sub>2</sub> enriched atmosphere on enzymes of the primary carbon metabolism of spruce trees. Plant Physiol Biochem 30: 541–547
- Wall, GW and Kimball BA (1993) Biological databases derived from free air carbon dioxide enrichment experiments. In: Schulze ED and Mooney HA (eds) Design and Execution of Experiments on CO<sub>2</sub> Enrichment, pp 329–351. Report No. 6, Ecosystems Research Report Series, Environmental Research Programme, Commission of the European Communities, Brussels
- Wall GW, Adam NR, Brooks TJ, Kimball BA, Pinter PJ Jr, LaMorte RL, Adamsen FJ, Hunsaker DJ, Wechsung G, Wechsung F, Grossman-Clarke S, Leavitt SW, Matthias AD and Webber AN (2000) Acclimation response of spring wheat in a free-air CO<sub>2</sub> enrichment (FACE) atmosphere with variable soil nitrogen regimes. 2. Net assimilation and stomatal conductance of leaves. Photosynth Res 66: 79–95 (this issue)
- Wechsung G, Wechsung F, Wall GW, Adamsen F, Kimball BA, Garcia RL, Pinter PJ Jr. and Kartschall TH 1995 Biomass and growth rate of a spring wheat root system grown in free-air  $\rm CO_2$  enrichment (FACE) and ample moisture. J Biogeogr 22: 623–634
- Webber AN, Long SP and Nie G-Y (1994) Effects of rising CO<sub>2</sub> concentration on expression of photosynthetic proteins. Photosynth Res 39: 413–426
- Wong SC (1979) Elevated atmospheric partial pressure of CO<sub>2</sub> and plant growth. I. Interactions of nitrogen nutrition and photosynthetic capacity in C<sub>3</sub> and C<sub>4</sub> plants. Oecologia: 68–74
- Wullschleger SD (1993) Biochemical limitations to carbon assimilation in  $C_3$  plants a retrospective analysis of the A/ $C_i$  curves from 109 species. J Exp Bot 44: 902–920
- Ziska LH, Hogan KP, Smith AP and Drake BG (1991) Growth and photosynthetic response of nine tropical species with long-term exposure to elevated carbon dioxide. Oecologia 86: 383–389